2016 JULY

07/01 Human practice

Court & Police office visiting detail

07/11 Human practice

Video Chatting with Israel iGEM Team

07/29 Human practice

Chung Shan Department of Laboratory Medicine visiting

07/30 Human practice

NCTU NTHU CGU iGEM meet up

07/04 Lab

Transformation of E. coli BL21 with LO, LO-GFP, GFP/pET-29b

07/05 Lab Web

1. To keep the original wiki code
2. Fill the tips
3. Culture bacteria of BL21carrying LO, LO-GFP, GFP/pET-29b
4. TA cloning procedure (PCR, cleanup, ligation)

07/06 Lab

1. IPTG induction of E. coli BL21 expressing LO, LO-GFP, GFP/pET-29b, pET-29bas control
2. Observing GFP expression under fluorescent microscope
3. Collect protein lysates
4. TA cloning continued: transformation
5. Freeze E. coli BL21bacteria of LO, LO-GFP, GFP/pET-29b, pET-29b as control
6. Gel extraction of AOX1, AOX2 DNA fragment amplified for cloning into pET-29b vectors

07/07 Lab

1. TA cloning continued: colony PCR
2. Discussion for Alcohol Monitoring System activities
3. Preparation of agarose gel
4. PCR with the templates from gel extraction (measure DNA conc. and dilute 10 times before PCR)

07/11 Lab

1. AOX1, 2 --> Gel extraction --> RE cut O/N
2. Repeat TA cloning
3. Culture BL21 for testing IPTG induction

07/12 Lab

1. Autoclave materials
2. Make stocks, Plasmid extraction, RE check for TA cloning
3. Ligation & Transformation for cloning AOX1, 2onto LO/pET-29b & pET029b4. Try conditions for IPTG induction

07/13 Lab

1. Prepare LB media & Kan Agar Plate
2. Install Microsoft Office on Laptop Dell
3. Call Boss Yu about BioTek & Gen5Software for help
4. Colony PCR: LO-/+ AOX1, 2/pET-29b
5. Sequencing: TA plasmids
6. Restriction enzyme cut for EV71-related gBlock fragment & GOX2016/07/19IPTG induction: test condition

07/21 Lab

1. Cleanup --> Ligation --> Transformation (GOX+pSB1C3)
2. Retransform: DH5alpha for #2-1 (LO-AOX1), BL21for AOX2, LO-AOX23. Prepare materials for SDS-PAGE & WB

07/22 Lab

1. LO-AOX2/pET-29b: Plasmid extraction --> RE check
2. GOX/pSB1C3: Colony PCR

07/25 Lab

1. Colony PCR --> Run gel
2. Prepare Lysis buffer, SDS-PAGE & WB materials
3. Prepare Amp agar plates4. Autoclave 125 ml flask

07/27 Lab

1. Freeze for stock: AOX2, LO-AOX2/pET-29b E. coli BL21
2. IPTG induction
3. Enzyme activity assay4. Store protein lysates

07/28 Lab

SDS-PAGE & WB2016/07/29SDS-PAGE & WB continued